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Ovine bladder smooth muscle cell morphology and adhesion on coverslips coated with or without proteins. Thirty minutes prior to cell seeding, serum free DMBM and DMEM supplemented with 10% FBS and 1% P/S were placed onto the protein coated substrates and reference substrates, respectively.

5 Bladder smooth muscle cells were enzymatically lifted from polystyrene tissue culture dishes using 1 ml of PET and resuspended in serum free DMEM. Cells were seeded ($3,500 \text{ cells/cm}^2$) onto each substrate and incubated under standard cell culture conditions for 4 h.

At the end of each time period, non-adherent cells were removed by
10 gentle rinsing in PBS. Cells adherent to the substrates were fixed with 10% formalin, stained with Coomassie Brilliant Blue for 10 minutes, and rinsed with PBS. Cell morphology was examined using light microscopy (Nikon Microphot-FXA; Nikon, Japan) and digital imaging (Flashbus FBG 4.2; Integral Technologies, Inc.). In addition, the number of cells in each of five random fields per substrate were counted
15 using light microscopy (Olympus CR2; Olympus, Germany), averaged, and reported as cell density (cells/cm^2). All experiments were run in triplicate and repeated a minimum of three separate times.

Bladder smooth muscle cells adherent on glass coverslips coated with collagen type IV in the presence of serum free DMEM exhibited superior spread
20 morphology compared to those adherent on glass coverslips coated with vitronectin and collagen type III, which exhibited superior morphology to those adherent on the albumin-coated control coverslips.

In addition, collagen type IV enhanced bladder smooth muscle cell adhesion compared to the albumin control coverslips. Cell adhesion was significantly
25 ($p < 0.05$) increased on coverslips coated with gelatin, fibronectin, vitronectin, laminin, and collagen type I, and was significantly ($p < 0.01$) increased on glass coverslips coated with collagen type IV, after 4 h of incubation. The adherent cell density on glass coverslips coated with collagen type IV was 762%, 324%, 148%, 132%, 96%, 86%, and 58% higher than on glass coverslips coated with laminin,
30 collagen type I, albumin, fibronectin, collagen type III, vitronectin, and gelatin, respectively.

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L3	0	"6030635".pn. and gelatin	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 07:53
L4	2	"20030143283".pn.	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 07:55
L5	2	"20020173033".pn.	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:02
L6	2	"6344367".pn.	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 08:02
L7	2	"6689374".pn.	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 08:02
L8	652542	(nanosized or nano) ith implant	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:02
L9	192	(nanosized or nano) with implant	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:02
L10	9	(nanosized or nano) with implant with film	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:04
L11	43541	polylactic or polyglycolic or polyetherurethane or polycaprolactone	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:05
L12	22	l9 and l11	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:05

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L14	2	"6270347".pn.	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:18
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L16	267064	l11 an dl15	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:20
L17	170	l11 and l15	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:20
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L19	163	cell and l17	US-PGPUB; USPAT; USOCR; EPO; JPO; DERWENT	OR	ON	2007/05/23 09:21